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CONCERNING A FILING UNDER 35 U.S.C. 371

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09/700713

INTERNATIONAL APPLICATION NO.
PCT/JP00/01764INTERNATIONAL FILING DATE
2000 March 23PRIORITY DATE CLAIMED
26 March 1999

TITLE OF INVENTION: ADDITIVES FOR CRUSTACEAN OR FISH FEEDS AND FEEDS

APPLICANT(S) FOR DO/EO/US: Genichiro SOMA, Yukinori TAKAHASHI, Denichi MIZUNO

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
(THE BASIC FILING FEE IS ATTACHED)
 2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
 3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT articles 22 and 39(1).
 4. ☒ A proper demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
 5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
 6. ☒ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
 7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
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 8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
 9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
 10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
- Items 11. to 16. below concern other document(s) or information included:
11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
 12. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
 13. ☒ A FIRST preliminary amendment.
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09/700713

INTERNATIONAL APPLICATION
NO. PCT/JP00/01764

ATTORNEY DOCKET NO. 101149-00008

DATE: November 27, 2000

17. XX The following fees are submitted:**Basic National Fee (37 CFR 1.492(a)(1)-(5)):**

Search Report has been prepared by the EPO or JPO.....\$860.00
 International preliminary examination fee paid to USPTO (37 CFR 1.492).....\$690.00
 No international preliminary examination fee paid to USPTO (37 CFR 1.492) but
 international search fee paid to USPTO (37 CFR 1.492(a)(2)).....\$710.00
 Neither international preliminary examination fee (37 CFR 1.492) or international
 search fee (37 CFR 1.492(a)(3)) paid to USPTO.....\$1,000.00
 International preliminary examination fee paid to USPTO (37 CFR 1.492) and all
 claims satisfied provisions of PCT Article 33(1)-(4).....\$ 100.00

CALCULATIONS

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Claims	Number Filed	Number Extra	Rate	
Total Claims	16 - 20 =	00	X \$ 18.00	\$00
Independent Claims	01 - 3 =	00	X \$ 80.00	\$00
Multiple dependent claim(s) (if applicable)			+ \$270.00	\$00

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 months from the earliest claimed priority date (37 CFR 1.492(f)).

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NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

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09/700713

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:

Genichiro SOMA et al

Serial No.: New Application

Filed: November 27, 2000

For: ADDITIVES FOR CRUSTACEAN OR FISH FEEDS AND FEEDS

PRELIMINARY AMENDMENT

Commissioner of Patents
Washington, D.C. 20231

November 27, 2000

Sir:

Prior to calculation of the filing fee and prior to the examination of this application, please amend the above-identified application as follows:

IN THE SPECIFICATION:

Page 1, before the first line of the specification insert --This application is the National Stage Application of PCT/JP00/01764 filed March 23, 2000.--

Page 21, line 15, delete "5 groups" and insert therefor --7 groups--.

Page 26, Table 8, after Group 4, delete "low" and insert therefor --high--.

after Group 5, delete "low" and insert therefor --high--.

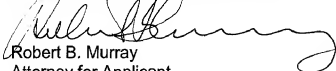
Page 27, line 23, delete "shrimp's" and insert therefor --yellowtail's--.

REMARKS

The above amendments have been made to put the application in better condition for examination.

In the event that any fees are due in connection with this paper, please charge our
Deposit Account No. 01-2300.

Respectfully submitted,
ARENT FOX KINTNER PLOTKIN & KAHN PLLC


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SPECIFICATION

ADDITIVES FOR CRUSTACEAN OR FISH FEEDS AND FEEDS

5 TECHNICAL FIELD

The present invention relates to a feedstuff additive for crustaceans or fishes, and a feed containing the feedstuff additive, and more particularly to a feedstuff additive which shows significant effects of activating immunity and preventing infection and to a feed containing the same in a suitable proportion.

BACKGROUND ART

Recent years have seen development of aquiculture of crustaceans and fishes. Attendant on the development is a great economical damage in the culture industry due to outbreaks of bacterial or viral diseases of crustaceans and fishes. Diseases of crustaceans and fishes often occurring include acute viremia of kuruma prawns (*Penaeus japonicus*), vibriosis thereof, pseudotuberculosis of yellowtails, enterococcus diseases thereof, cold-water disease of sweet fishes (ayu), *Pseudomonas* diseases thereof, iridovirus diseases of red sea breams, *Seriola dumerili*, yellowtails or the like which have economically damaged the culture industry. Of these diseases, bacterial diseases have been treated with antibiotics or synthetic antibacterial agents as a curative agent. However, with the advent of antibiotic-resistant bacteria, satisfactory curative effects have not been achieved. Further, a problem of

public health hazards has been raised because of the medicinal agent remaining in crustaceans and fishes. Consequently, there is a strong demand for preventive measures not depending on chemotherapy. On the other hand, vaccines and curative agents have not been developed against viral diseases of crustaceans and fishes and viral diseases still often occur.

The use of polysaccharides is already known to immunopotentiate crustaceans and fishes and to prevent infectious diseases thereof. These polysaccharides include, for example, peptidoglycan derived from *Bifidobacterium thermophilum* (Patent No.2547371), cell wall-forming component of gram-positive bacteria like bacteria of genus *Bacillus* (JP-B-3-173826) and β -1,3-glucan derived from *Schizophyllum commune* (JP-B-6-65649). It was already reported that high molecular weight lipopolysaccharides activate the immune function of fishes and animals (Salati, F. and R. Kusuda, Society Journal, Japanese Society of Science of Fisheries, vol.53, pp.201 to 204, 1987 and Odean, M.J. et al., Infection and Immunity, vol.58, pp.427 to 432, 1990).

On the other hand, the low molecular weight lipopolysaccharide of the present invention (hereinafter referred to as "low molecular weight LPS") is different in basic structure and components from the peptidoglycan derived from gram-positive bacteria, cell wall-forming component and β -1,3-glucan derived from a mushroom. The low molecular weight LPS of the invention comprises three components, i.e. a specific lipid A, an oligosaccharide with covalent bond therewith called R core and O specific polysaccharide. The low

molecular weight LPS of the invention is known as an immunopotentiator for animals because of its ability to increase the tumor necrosis factor (TNF)-producing effect, but is not known at all to have an activity of preventing
5 infectious diseases of crustaceans and fishes. The high molecular weight lipopolysaccharides (LPSSs) used in the researches heretofore reported are those with a markedly high molecular weight as high as 1 million to 10 millions and are of high toxicity. Consequently, when applied to crustaceans and
10 fishes for a long period, such high molecular weight LPS is unable to activate the immune function all the time. The above-mentioned known substances have a high molecular weight and need to be orally administered in a large quantity because of their poor absorption through the intestinal tract.
15 Consequently, a long-period intake of them frequently results in impairment of immune function.

As described above, a variety of infectious diseases often occur in crustaceans and fishes. Some of these diseases are lethal and may result in great economic damage. The
20 background to be noted is that the immune function of crustaceans and fishes is deteriorated because they are bred in an overcrowded area under a limited environment. Various substances were used to reactivate their impaired immune system. On the other hand, crustaceans have no ability to produce an
25 antibody nor lymphocyte, neutrophile or basophile as found in a vertebrate. Fishes have a limited ability to produce an antibody and its production of antibody is greatly affected by the temperature of water because they are cold-blooded animals

so that such immune system is not sufficiently functioned. In other words, substantial difference exists in defensive mechanism between these oceanic organisms and mammals (Fish Pathology, 30(2), 141-150, June in 1995). Consequently some of the substances are not usable in-situ in breeding oceanic organisms because of high toxicity like conventional LPSSs, and most of them are impaired in the immune system by intake of the LPSSs for a prolonged period.

An object of the present invention is to provide a safe feedstuff additive for culture or breeding of crustaceans and fishes, the feedstuff additive being capable of preventing infectious diseases even in a small amount by properly activating their intrinsic immune function, and being free from problems of public health hazards such as the feedstuff additive remaining in crustaceans and fishes.

DISCLOSURE OF THE INVENTION

The present invention provides a feedstuff additive for crustaceans and fishes, characterized in that it is prepared from gram-negative bacteria, that it has a molecular weight of 5000 ± 2000 as measured by SDS-PAGE method using a protein marker, that it is substantially free of high molecular weight lipopolysaccharide, and that it contains a low molecular weight lipopolysaccharide as an effective component and that it is capable of activating immunity or preventing infection in crustaceans or fishes; and a feed for crustaceans or fishes which feed is characterized in that it contains the feedstuff additive.

The present invention also provides a feedstuff additive for crustaceans or fishes comprising the low molecular weight lipopolysaccharide and a carrier acceptable for crustaceans and fishes.

5 The present invention also provides use of the low molecular weight lipopolysaccharide for the preparation of a feedstuff additive for crustaceans or fishes.

The present invention also provides a method of activating immunity or preventing infection in crustaceans and fishes comprising administering an effective amount of the low molecular weight lipopolysaccharide to crustaceans or fishes.

10 The present invention also provides an agent for preventing the perish of crustaceans or fishes comprising the low molecular weight lipopolysaccharide as an effective component.

15 The present invention also provides an agent for preventing the perish of crustaceans or fishes comprising the low molecular weight lipopolysaccharide and a carrier acceptable for crustaceans and fishes.

20 The present invention also provides use of the low molecular weight lipopolysaccharide of for the preparation of an agent for preventing the perish of crustaceans or fishes.

25 The present invention also provides a method of preventing the perish of crustaceans or fishes comprising administering an effective amount of the low molecular weight lipopolysacchride to crustaceans or fishes.

The present invention also provides a feedstuff additive, wherein the gram-negative bacteria are those pertaining to

genus Pantoea.

The present invention also provides a feedstuff additive, wherein the gram-negative bacteria are Pantoea agglomerans.

The present invention also provides a feed for
5 crustaceans or fishes comprising the feedstuff additive.

The present invention also provides a feed for crustaceans or fishes comprising the agent for preventing the perish.

The present invention also provides a method of breeding
10 crustaceans or fishes comprising administering the feed to crustaceans or fishes.

The feedstuff additive of the invention is prepared from gram-negative bacteria by purification, e.g. according to the method disclosed in JP-A-8-198902. The present inventors
15 prepared a feed containing a low molecular weight LPS having a molecular weight of 5000 ± 2000 . When the feed was supplied to crustaceans and fishes, it was found that the feed prevented viral or bacterial infectious diseases and protected them against disease by activation of the intrinsic immune function.
20 The present invention was accomplished based on this finding.

The low molecular weight LPS of the present invention is, as described above, a lipopolysaccharide having a molecular weight of 5000 ± 2000 which is prepared from gram-negative bacteria, e.g. according to the method disclosed in JP-A-8-
25 198902. The LPS of this invention is characterized in that the LPS is pronouncedly safer for crustaceans or fishes and can produce a significantly higher effect of activating immunity and a higher effect of preventing infection and disease than

conventional LPSs (with a molecular weight of 1 million to 10 millions).

In the present invention, the term "substantially free of high molecular weight lipopolysaccharide" means "not containing
5 lipopolysaccharide having a molecular weight of at least 8,000".

The gram-negative bacteria for use in the invention include, for example, those pertaining to genera Pantoea, Salmonella, Aeromonas, Serratia and Enterobacter, and further include those described in JP-A-4-99481. Among useful gram-
10 negative bacteria, those of Pantoea are preferred and those of Pantoea agglomerans are more preferred.

The low molecular weight LPS of the present invention can be prepared by a method comprising incubating gram-negative bacteria or the like in the conventional manner, collecting the
15 cultured bacteria from the culture medium, extracting the collected bacteria by conventional methods, such as hot phenol method (edited by O. Westphal, Methods in Carbohydrate Chemistry, vol. 5, p.83, Academic Press, 1965) and purifying the extract with an anion exchange resin. More specifically,
20 the method comprises suspending bacteria in distilled water, adding the suspension to a mixture of distilled water and an equal volume of hot phenol, stirring the mixture, centrifuging the mixture to recover the aqueous layer, dialyzing the aqueous layer to remove the phenol, concentrating the aqueous layer by
25 ultrafiltration to obtain crude LPS fractions, purifying the fractions by conventional anion exchange chromatography (e.g. using mono Q-Sepharose or Q-Sepharose) and desalting the same in the conventional manner.

The purified LPS thus obtained is substantially identical with the LPSs having a molecular weight of about 5,000 to about 6,000 as disclosed in JP-A-4-187640, JP-A-4-49240, JP-A-4-99481 and JP-A-5-155778. The purified LPS is subjected to gel

5 filtration in the presence of a surface-active agent such as sodium deoxycholate to recover only low molecular weight LPS-containing fractions, whereby only a highly purified low molecular weight LPS is obtained by removal of the high molecular weight LPS from the fractions. The procedure of gel
10 filtration in the presence of a surface-active agent is carried out to more highly purify the LPSs having a molecular weight of about 5,000 to about 6,000 which are disclosed in JP-A-4-187640, JP-A-4-49240 and JP-A-5-155778, whereby the high molecular weight LPS is completely removed from the fractions.

15 The term "crustaceans" used herein refers to all of lobsters, shrimps or prawns such as kuruma prawn (*Penaeus japonicus*), ushi prawn (*Penaeus monodon*), Yellow Sea prawn (*Penaeus chinensis*) and banana prawn (*Penaeus merguensis*), and all of crabs such as *Portunus trituberculatus* and Chinese
20 mitten crab, preferably lobsters, shrimps or prawns, more preferably prawns. The term "fishes" used herein include all of fishes such as yellowtail, globefish, real sea bream, flatfish, eel and rainbow trout. The infectious diseases referred to herein include acute viremia of crustaceans, their
25 vivrio diseases, parasitosis such as *Bpistylis* sp., *Zoothamnium* sp. or mycosis such as *Lagenidium* sp., *Siropidium* sp.; iridovirus infectious diseases of fishes, their rhabdovirus diseases, neuronecrosis, infectious hemopoietic organ necrosis,

pseudotuberculosis, streptococcic diseases, enterococcus diseases, vibrio diseases, cold-water disease, Pseudomonas diseases, gliding-bacteria diseases and Saprolegnia diseases, and all of infectious diseases caused by viruses, mycoplasmas, bacteria, fungi and parasites among which the feedstuff additive and feed of the invention can be more effectively used for viremia of crustaceans, and fishes' diseases such as streptococcic diseases, enterococcus diseases and vibrio diseases.

The low molecular weight LPS of the present invention can be used as a feed additive for crustaceans and fishes, and for this purpose, may be used as it is or as mixed with conventional carriers, stabilizers and the like and optionally with vitamins, amino acids, minerals and like nutrients, antioxidants, antibiotics, antibacterial agents and other additives. The feed additive is prepared in a suitable form such as powders, granules, pellets or suspensions. The feed additive may be supplied to crustaceans or fishes, alone or in mixture with a feed. For prevention of diseases, the feed additive may be supplied together with the feed at all times or at a latter half of feeding time.

The feeds of the present invention are not specifically limited but can be any of powdery feeds, solid feeds, moist pellet feeds, dry pellet feeds, extruder pellet feeds and live baits.

The proportion of the low molecular weight LPS in the feed of the invention can be selected from a wide range and is preferably 0.000001 to 0.001% by weight, more preferably

0.00002 to 0.00005% by weight to which its proportion is not limited. The amount of the low molecular weight LPS to be used can be suitably determined. For example, the LPS is applied at a daily dose of 1 to 100 μ g, preferably 10 to 20 μ g, per kilogram of the body weight of crustaceans or fishes to which, however, the dose is not limited.

BEST MODE OF CARRYING OUT THE INVENTION

The present invention will be described in detail with reference to the following Examples to which, however, the invention is not limited. Low molecular weight LPS used in Examples is LPS having a molecular weight of about 5,000, and high molecular weight LPS is LPS having a molecular weight of about 8,000 to 50,000.

Reference Example 1 (Preparation of low molecular weight LPS)

A 10 g quantity of tryptone (product of DIFCO CO.), 5 g of yeast extract (product of DIFCO CO.) and 10 g of NaCl (product of WAKO PURE CHEMICAL INDUSTRIES, LTD., special grade) were added to 1 liter of distilled water. The suspension was adjusted to a pH of 7.5 with NaOH and was sterilized in an autoclave. A single colony was separated from *Pantoea agglomerance*-carrying bacteria maintained at -80°C and was inoculated in a 500 ml-vol. Sakaguchi flask holding 100 ml of a culture medium containing sterile glucose (product of WAKO PURE CHEMICAL INDUSTRIES, LTD., special grade) at a proportion of 0.1% (hereinafter referred to as L-broth medium). Then the cells were subjected to shake culture at 35°C overnight. The cultured cells were inoculated in its entirety in a 3 liter-vol.

Sakaguchi flask holding 1,000 ml of L-broth medium and were further cultivated in the same manner as above.

The cultured cells were inoculated in a 10-liter vol. desk fermenter (product of MARUBISHI BIOENGI CO.) holding 7
5 liters of L-broth medium and were subjected to aeration culture under the same conditions. The cells were collected to recover about 70 g of wet bacteria and were freeze-stored. About 70 g of freeze-stored cells were suspended in 500 ml of distilled water. A 500-ml quantity of 90% hot phenol was added to the
10 suspension. The mixture was stirred at 65 to 70°C for 20 minutes and was cooled. The mixture was centrifuged at 10,000 G and 4°C for 20 minutes to recover the aqueous layer. The phenol layer was treated in the same manner as above. Then the two aqueous layers thus obtained were combined and dialyzed
15 overnight to remove the phenol. The inner solution was concentrated by ultrafiltration in a 2 atom. nitrogen gas using an ultrafiltration device (product of ADVANTEC TOYO CO., K-200) with a membrane filter by cutting off molecular weight 200,000.

The lyophilized product of crude LPS thus obtained was
20 dissolved in distilled water, the filter was sterilized, a buffer was added, and the solution was subjected to anion exchange chromatography (product of PHARMACIA Co., Q-Sepharose first flow). The specimen solution was passed through the column using a buffer containing 10 mM Tris-HCl (pH 7.5) and 10
25 mM NaCl to elute a limulus active fraction with 200 to 400 mM NaCl/10 mM Tris-HCl (pH 7.5). The eluate was subjected to ultrafiltration under the same conditions as above for desalting and concentration and was lyophilized to obtain about

300 mg of purified LPS from about 70 g of wet bacteria.

The obtained purified LPS (100 mg) was dissolved in a solubilizing buffer [comprising 3% sodium deoxycholate (product of WAKO PURE CHEMICAL INDUSTRIES LTD.), 0.2 M sodium chloride, 5 mM EDTA-2Na and 20 mM Tris-hydrochloric acid, pH 8.3]. The purified LPS solution (20 ml) was gently placed over Sephacryl S-200 HR column (product of PHARMACIA CO.). Then, 800 ml (50 hours) of the solution was eluted with an eluting buffer [comprising 0.25% sodium deoxycholate (product of WAKO PURE CHEMICAL INDUSTRIES LTD.), 0.2 M sodium chloride, 5 mM EDTA and 10 mM Tris hydrochoric acid, pH 8.3] at a flow velocity of 16 ml/hr.

The obtained eluate was fractionated by a fraction collector (product of ADVANTEC CO., trade name SF 2120) under control of flow velocity using a perista-pump PI (product of PHARMACIA CO.). A first 240-ml portion (24- fraction portion) was cast away. Thereafter the residue was fractionated into 80 fractions at 10 ml/fraction. The saccharide in the eluted fractions was quantitatively determined using the base solution or diluted solution by phenol/sulfuric acid method (Sakuzo FUKUI, "Method of Quantitative Determination of Reducing Sugar", 2nd ed., pp. 50 to 52, Gakkai Shuppan Center, 1990) to check the elution state. The fraction pattern of LPS was investigated by SDS-PAGE method using 0.5 ml of each of fractions 37 to 55 among the fractions presumably having LPS (fractions 30 to 60).

The result of investigation demonstrates that the fractions 45 to 55 contained only low molecular weight LPS

(m.w. about 5000) and that fractions 37 to 44 contained both low molecular weight LPS and high molecular weight LPS. The low molecular weight LPS fractions of fractions 45 to 55 were further purified as follows.

5 The fractions was mixed, lyophilized and suspended in ethanol. The suspension was centrifuged to remove the deoxycholic acid soluble in ethanol and to recover a low molecular weight LPS in insoluble fractions. The ethanol treatment of the low molecular weight LPS fractions was further repeated twice, followed by removal of deoxycholic acid. The 10 obtained LPS was suspended in 70% ethanol again, and the buffer component was removed by centrifugation. The same procedure was repeated three times for recovery of low molecular weight LPS in the insoluble fractions, followed by lyophilization, 15 whereby about 20 mg of purified low molecular weight LPS was produced.

Example 1 (Safety of low molecular weight LPS in crustaceans)

Kuruma prawns having an average weight of 20 g were divided into 5 groups of each 20 prawns. The low molecular 20 weight LPS of the present invention was intramuscularly administered to the third abdominal segment of prawns in Groups 1 and 2 at a dose of 50 mg and 100 mg, respectively per kilogram of the prawn's weight. On the other hand, a conventional high molecular weight LPS (derived from E. coli, E. 25 coli 0111 manufactured by DIFCO CO.) was intramuscularly administered to the third abdominal segment of prawns in Groups 3 and 4 at a dose of 10 mg and 20 mg, respectively per kilogram of the prawn's weight. Group 5 received a physiological saline

free of LPS. The life or death of prawns up to 120 hours after administration was checked to determine a mortality. The results are shown in Table 1.

5

Table 1

<u>Group</u>	<u>number of perish</u> <u>/ number tested</u>	<u>mortality</u> <u>(%)</u>
Group 1 low MW LPS 50 mg/kg	0/20	0
Group 2 low MW LPS 100 mg/kg	0/20	0
Group 3 high MW LPS 10 mg/kg	13/20	65
Group 4 high MW LPS 20 mg/kg	20/20	100
Group 5 physiological saline	0/20	0

As apparent from Table 1, a mortality of prawns in the groups receiving 10 mg or 20 mg of high molecular weight LPS was 65 or 100%, respectively, whereas no prawn died in the groups receiving 50 mg and 100 mg of low molecular weight LPS. It is clear from the above data that low molecular weight LPSs are significantly safe for prawns as compared with conventional high molecular weight LPSs.

15 Example 2 (Safety of low molecular weight LPS in fishes)

Black carps having an average weight of 85 g were divided into 3 groups of each 40 carps. The low molecular weight LPS of the present invention was intramuscularly administered to the dorsal region of black carps in Group 1 at a dose of 100 mg per kilogram of the carp's weight. On the other hand, a conventional high molecular weight LPS (trade name E. coli 0111 manufactured by DIFCO CO.) was intramuscularly administered to

20

the dorsal region of black carps in Group 2 at a dose of 20 mg per kilogram of the carp's weight. Group 3 received a physiological saline free of LPS. The life or death of black carps up to 120 hours after administration was checked to determine a mortality. The results are shown in Table 2.

Table 2

<u>Group</u>	<u>number of perish</u> <u>/ number tested</u>	<u>mortality</u> <u>(%)</u>
Group 1 low MW LPS 100 mg/kg	0/40	0
Group 2 high MW LPS 20 mg/kg	34/40	85
Group 3 physiological saline	0/40	0

As apparent from Table 2, a mortality of black carps was 85% in the group receiving 20 mg of high molecular weight LPS, whereas no black carp died in the group receiving 100 mg of low molecular weight LPS. It is clear from the above data that the low molecular weight LPS of the present invention is significantly safe for fishes as compared with conventional high molecular weight LPS.

Example 3 (Activity of activating phagocytosis in hemocyte of crustaceans)

Kuruma prawns having an average weight of 20 g were divided into 6 groups of each 20 prawns. Groups 1, 2 and 3 received the low molecular weight LPSs of the present invention as admixed with feeds at a daily dose of 20, 40 and 100 μ g, respectively per kilogram of prawn's weight. On the other hand, Group 4 received a high molecular weight LPS as admixed with a

feed at a daily dose of 100 μg , and Group 5 received the same at a daily dose of 1000 μg , per kilogram of prawn's weight. The feeds were given for 7 days. Group 6 was given a feed free of LPS. On day 0, day 1, day 5 and day 7 after supply of the feeds, the blood was collected from the thorax recess of prawns using a syringe holding a K-199 culture medium containing L-cystein as an anticoagulant. Hemocyte cells were obtained by centrifugation. The obtained cells (1×10^5 cells per microliter of the suspension) were mixed with 1×10^8 latex beads ($1.986 \mu\text{m}$ in diameter) and were reacted at 25°C for 30 minutes. After fixing the reaction mixture with glutaraldehyde, it was air-dried. Then the mixture was subjected to giemsa staining and was fixed to a slide glass with Eukitt. The same procedure was repeated to obtain five samples per prawn. The hemocyte cells (200 cells per sample) were observed at random under an epi-fluorescent microscope to determine the phagocytosis ratio of latex beads in hemocyte and the number of latex beads phagocytized into one cell of hemocyte. Then the phagocytosis index was calculated by the following equation.

Phagocytosis ratio = $\left[\frac{\text{number of hemocyte cells taking beads}}{\text{total number of hemocyte cells observed}} \right] \times 100$.

Average number of beads taken by hemocyte cells = $\frac{\text{number of beads taken by hemocyte cells}}{\text{number of hemocyte cells taking beads}}$.

Phagocytosis index = $\left[\frac{\text{number of hemocyte cells taking beads}}{\text{total number of hemocyte cells observed}} \right] \times \left[\frac{\text{number of beads taken by hemocyte cells}}{\text{total number of hemocyte cells observed}} \right] \times 100$.

Test results: The biophylaxis of crustaceans involves a cell

factor and a liquid factor. The phagocytosis of foreign particles in hemocyte is deeply concerned with the former. When the phagocytosis of foreign particles in prawn's hemocyte is assessed, it is clarified whether the defensive mechanism of prawns is activated. [Yukinori TAKAHASHI et al, Research of Fish Diseases, 30 (2), pp.141 to 150, (1995)]. In view of said theory, the phagocytosis index was determined on day 0, day 1, day 5 and day 7 after supply of feeds for the groups receiving high molecular weight LPSs and the groups receiving the low molecular weight LPSs. The results were tabulated in Table 3.

Table 3

Group	Phagocytosis index of hemocyte	
	0	1 day
Group 1 low MW LPS 20 $\mu\text{g/kg}$	0.9 ± 0.18	2.1 ± 0.61 *2
Group 2 low MW LPS 40 $\mu\text{g/kg}$	0.9 ± 0.18	3.3 ± 1.16 *2
Group 3 low MW LPS 100 $\mu\text{g/kg}$	0.9 ± 0.18	3.8 ± 1.00 *2
Group 4 high MW LPS 100 $\mu\text{g/kg}$	0.9 ± 0.18	0.7 ± 0.31
Group 5 high MW LPS 1000 $\mu\text{g/kg}$	0.9 ± 0.18	1.1 ± 0.63
Group 6 feed free of LPS	0.9 ± 0.18	0.5 ± 0.24

Table 3 (continued)

Group	Phagocytosis index of hemocyte	
	5 days	7 days
Group 1 low MW LPS 20 μ g/kg	3.2 \pm 0.71 *2	8.4 \pm 1.37 *2
Group 2 low MW LPS 40 μ g/kg	4.5 \pm 0.75 *2	3.7 \pm 1.02 *2
Group 3 low MW LPS 100 μ g/kg	3.1 \pm 0.94 *2	2.8 \pm 0.70 *1
Group 4 high MW LPS 100 μ g/kg	0.7 \pm 0.82	1.2 \pm 0.44
Group 5 high MW LPS 1000 μ g/kg	2.1 \pm 0.58 *1	2.9 \pm 0.68 *1
Group 6 feed free of LPS	0.7 \pm 0.5	1.1 \pm 0.56

*1: Significant difference between this group and Group 6
(P<0.05)

*2: Significant difference between this group and Group 6
(P<0.01)

As apparent from Table 3, the groups receiving the low molecular weight LPSs (present invention) showed a higher phagocytosis index in hemocyte of prawns than Group 6 and a significant difference in this index from Group 6 (P<0.01, P<0.05). The group receiving 100 μ g of conventional high molecular weight LPS was unable to increase the phagocytosis index in hemocyte of prawns after 1, 5 and 7 days. However, the group receiving 1000 μ g of conventional high molecular weight LPS showed a significantly higher phagocytosis index in hemocyte of prawns (P<0.05) than Group 6 after 5 and 7 days. The above data show that the low molecular weight LPSs of the present invention can activate the defensive mechanism such as phagocytosis in hemocyte of prawns even when used in an

extremely smaller amount than the high molecular weight LPSs.

Example 4 (Activity of activating phenol oxidase in hemocyte of crustaceans)

Kuruma prawns having an average weight of 20 g were
5 divided into 6 groups of each 20 prawns. Groups 1, 2 and 3
received the low molecular weight LPSs of the present invention
as admixed with feeds at a daily dose of 20, 40 and 100 μg ,
respectively per kilogram of prawn's weight. Group 4 received
a high molecular weight LPS as admixed with a feed at a daily
10 dose of 100 μg , and Group 5 received the same as admixed with
a feed at a daily dose of 1000 μg , per kilogram of prawn's
weight. The supply of the feeds continued for 7 days. Group 6
received a LPS-free feed. The blood was collected from the
thorax recess of prawns using a syringe holding a KHE culture
15 medium having EDTA on day 0, day 1, day 5 and day 7 after
supply of feeds. The collected blood was centrifuged to obtain
hemocyte cells. The obtained cells were suspended in a Ca-Mg
Hepes culture medium to a concentration of 1×10^6 cells/ml.
The cells were crushed by freeze resolution and supersonic
20 waves. The supernatant was separated off by centrifugation and
was filtered with a membrane filter. The obtained filtrate
(900 μl) was mixed with 100 μl of L-DOPA solution as a
substrate solution. Thereafter the mixture was reacted at a
temperature of 60°C for 60 minutes. Then the absorbance at 490
25 nm was measured by a spectrophotometer to assess a phenol
oxidase activity (PO activity).

Test results: The biophylaxis of crustaceans involves a cell

factor and a liquid factor. The PO activity in hemocyte is deeply concerned with the latter. Thus, it is clarified by assessment of PO activity whether the defensive mechanism of prawns is activated. The PO activity of prawns was determined on day 0, day 1, day 5 and day 7 after supply of feeds for the groups receiving the low molecular weight LPSs (present invention) and the groups receiving high molecular weight LPSs. The results were tabulated in Table 4.

Table 4

Group	PO activity (absorbance 490nm)			
	0	1 day	5 days	7 days
Group 1 low MW LPS 20 μ g/kg	0.092	0.105	0.199 *1	0.405 *2
Group 2 low MW LPS 40 μ g/kg	0.092	0.115	0.201 *1	0.325 *2
Group 3 low MW LPS 100 μ g/kg	0.092	0.166 *1	0.170 *1	0.292 *2
Group 4 high MW LPS 100 μ g/kg	0.092	0.093	0.124	0.138
Group 5 high MW LPS 1000 μ g/kg	0.092	0.104	0.197 *1	0.230 *1
Group 6 feed free of LPS	0.092	0.093	0.136	0.123

*1: significant difference between this group and Group 6
($P < 0.05$)

*2: significant difference between this group and Group 6
($P < 0.01$)

As apparent from Table 4, the groups receiving the low molecular weight LPSs (present invention) indicated a higher PO

activity than Group 6 and a significant difference in this activity from Group 6 ($P<0.01$, $P<0.05$). The group receiving 100 μg of conventional high molecular weight LPS did not increase in PO activity in hemocyte of prawns up to 7 days.

5 The group receiving 1000 μg of conventional high molecular weight LPS showed a significantly higher PO activity in hemocyte of prawns ($P<0.05$) than Group 6 after 5 and 7 days. The above data show that the low molecular weight LPSs of the present invention can activate the defensive mechanism such as
 10 PO activity in hemocyte of prawns even when used in an extremely smaller amount than the high molecular weight LPSs.

Example 5 (Effect of preventing acute viremia in kuruma prawns)

Kuruma prawns having an average weight of 14 g were
 15 divided into 5 groups of each 20 prawns. Groups 1, 2 and 3 received the low molecular weight LPSs of the present invention as admixed with feeds at a daily dose of 20, 40 and 100 μg , respectively per kilogram of prawn's weight. Group 4 received a high molecular weight LPS as admixed with a feed at a daily
 20 dose of 1000 μg , per kilogram of prawn's weight. Group 5 received peptidoglycan (PG) derived from Bifidobacterium thermophilum (Patent No.2547371) as admixed with a feed at a daily dose of 0.2mg (200 μg), per kilogram of prawn's weight. Group 6 received β -1,3-glucan (1,3-G) derived from
 25 Schizophyllum commune (JP-B-6-65649) as admixed with a feed at a daily dose of 50mg (50000 μg), per kilogram of prawn's weight. The supply of feeds continued for 18 days. Group 7 (control group) was given a LPS-free feed.

On day 8 after the start of supply of LPS, infection test was conducted using PRDV (penaeid rod-shaped DNA virus) as a pathogen inducing acute viremia in prawns. Carapaces were removed from the cephalothorax of three prawns which died of acute viremia. The intestine of prawns was crushed and homogenized in 40 ml of sterile seawater. The supernatant (10 ml) was separated off by centrifugation ($10,000 \times g$, 10 minutes, 4°C) and added to 20 liters of seawater. On day 8 after the start of supply of LPS, prawns were infected with acute viremia by immersion in the supernatant for 2 hours. The life or death of prawns was observed for 10 days after infection. The dead prawns were pathologically tested and examined by PCR (polymerase chain reaction) method to confirm whether the prawns died of infection with PRDV.

Test results: Tables 5 and 6 show the total number of dead prawns and a mortality after infection with PRDV in the groups receiving low molecular weight LPSs of the present invention, the group receiving a high molecular weight LPS and the group receiving a LPS-free feed.

Table 5

Group	Days after infection				
	1	2	3	4	5
Group 1 low MW LPS 20 μ g/kg	0	0	0	2*	3
Group 2 low MW LPS 40 μ g/kg	0	0	3	4	4
Group 3 low MW LPS 100 μ g/kg	1	1	3	3	4
Group 4 high MW LPS 1000 μ g/kg	1	1	6	6	6
Group 5 PG 0.2mg/kg	0	0	2	5	5
Group 6 1,3-G 50mg/kg	0	3	5	7	10
Group 7 feed free of LPS	2	4	13	14	15

* The number indicates the total number of dead prawns. (Other numbers show the same.)

Table 6

Group	Days after infection					Mortality
	6	7	8	9	10	
Group 1 low MW LPS 20 μ g/kg	3	3	4	4	4	20 ***
Group 2 low MW LPS 40 μ g/kg	6	6	6	7	7	35 ***
Group 3 low MW LPS 100 μ g/kg	5	6	8	8	8	40 ***
Group 4 high MW LPS 1000 μ g/kg	9	9	10	11	11	55 **
Group 5 PG 0.2mg/kg	7	8	8	8	10	50 **
Group 6 1,3-G 50mg/kg	10	11	11	12	12	60 **
Group 7 feed free of LPS	18	18	19	20	20	100

** : significant difference between this group and Group 7
($P < 0.05$)

*** : significant difference between this group and Group 7
($P < 0.01$)

5 All (100%) of prawns died in the control group receiving a LPS-free feed up to 9 days after infection with PRDV. On the other hand, 20%, 35% and 40% of prawns died in the groups receiving 20, 40 and 100 μg , respectively of low molecular weight LPS (present invention). In other words, a low
10 mortality resulted from these groups, and a significant difference ($P < 0.01$) exists between these groups and the control group. In contrast, 55% of prawns died in the group receiving 1000 μg of high molecular weight LPS, which means that more prawns died in this group than the groups receiving the low
15 molecular weight LPSs. The above data demonstrate that the low molecular weight LPSs of the present invention can prevent viral infection of prawns and that the low molecular weight LPSs are more efficacious than conventional high molecular weight LPSs.

20 Example 6 (Activity of activating immune function in fishes)

Yellowtails weighing 230 g on an average were divided into 6 groups of each 20 yellowtails. Groups 1, 2 and 3 received the low molecular weight LPSs of the present invention as admixed with moist pellets at a daily dose of 20, 40 and 100
25 μg , respectively per kilogram of yellowtail's weight. Group 4 received a high molecular weight LPS as admixed with moist pellets at a daily dose of 100 μg , and Group 5 received a high molecular weight LPS as admixed with moist pellets at a daily

dose of 1000 μ g, per kilogram of yellowtail's weight. The feeds were given for 7 days. Group 6 received LPS-free moist pellets. On day 0, day 1, day 5 and day 7 after supply of feeds, a head kidney was excised from 5 yellowtails. Then hemocyte cells were separated in a plastic petri dish holding a 0.25% NaCl-containing RPMI-1640-HAH culture medium. The cells were passed through a cell filter to give a cell suspension. The suspension was placed over a discontinuous Percoll density gradient. Thereafter a leukocyte layer was formed by centrifugation (1600 rpm., at 4°C for 20 minutes).

The leukocyte layer was collected and was subjected to centrifugal washing after which the cells were suspended in a 10% FBS (fetal bovine serum)-containing 0.25% NaCl-including RPMI-1640-H culture medium. The number of leukocyte cells in the suspension was adjusted to 1×10^6 cells/ml. The leukocyte suspension (500 μ l) and 500 μ l of a suspension (1×10^8 cells/ml) of yeast opsonized with serum of yellowtail were placed into a silicone-treated glass test tube and were incubated at 25°C for 60 minutes with stirring every 10 minutes. After incubation, 5 smears per yellowtail were produced, subjected to Wright's staining and enclosed with Eukitt. The hemocyte cells (200 cells per smear) were observed at random under an optical microscope. Then the number of yeast cells phagocytized into leukocyte was counted. The phagocytosis index was given by the same equation as in Example 3. The results are shown in Tables 7 and 8.

Table 7

Group	Phagocytosis index of leukocyte	
	0	1 day
Group 1 low MW LPS 20 $\mu\text{g/kg}$	7.3 ± 2.30	12.7 ± 2.65 *1
Group 2 low MW LPS 40 $\mu\text{g/kg}$	7.3 ± 2.30	17.9 ± 3.99 *2
Group 3 low MW LPS 100 $\mu\text{g/kg}$	7.3 ± 2.30	18.6 ± 4.12 *2
Group 4 high MW LPS 100 $\mu\text{g/kg}$	7.3 ± 2.30	6.3 ± 2.24
Group 5 high MW LPS 1000 $\mu\text{g/kg}$	7.3 ± 2.30	8.2 ± 2.18
Group 6 feed free of LPS	7.3 ± 2.30	6.6 ± 1.19

*1: significant difference between this group and Group 6

($P < 0.05$)

*2: significant difference between this group and Group 6

($P < 0.01$)

Table 8

Group	Phagocytosis index of leukocyte	
	5 days	7 days
Group 1 low MW LPS 20 $\mu\text{g/kg}$	39.2 ± 2.54 *2	52.7 ± 4.08 *2
Group 2 low MW LPS 40 $\mu\text{g/kg}$	37.4 ± 4.28 *2	37.0 ± 3.11 *2
Group 3 low MW LPS 100 $\mu\text{g/kg}$	42.6 ± 5.35 *2	36.5 ± 4.32 *1
Group 4 low MW LPS 100 $\mu\text{g/kg}$	11.2 ± 3.05	10.6 ± 2.96
Group 5 low MW LPS 1000 $\mu\text{g/kg}$	22.7 ± 3.16 *1	31.8 ± 3.52 *1
Group 6 feed free of LPS	9.0 ± 2.04	7.7 ± 1.73

*1: significant difference between this group and Group 6

($P < 0.05$)

**2: significant difference between this group and Group 6

($P < 0.01$)

As apparent from Tables 7 and 8, any groups of yellowtails receiving the low molecular weight LPSs (present invention) indicated a higher phagocytosis index in leukocyte of yellowtails than Group 6 and a significant difference ($P < 0.01$, $P < 0.05$) in this index from Group 6. However, the group receiving 100 μg of conventional high molecular weight LPS did not increase the phagocytosis index in leukocyte of yellowtails after 7 days. The group receiving 1000 μg of conventional high molecular weight LPS showed a significantly higher phagocytosis index ($P < 0.01$) in leukocyte of yellowtails than Group 6 after 5 days. The above data show that the low molecular weight LPSs of the present invention can activate the immune system of fishes such as phagocytosis in leukocyte in an extremely smaller amount than conventional high molecular weight LPSs.

Example 7 (Effect of preventing enterococcus disease in yellowtails)

Yellowtails weighing 63 g on an average were divided into 5 groups of each 30 yellowtails. Groups 1, 2 and 3 received the low molecular weight LPSs of the present invention as admixed with moist pellets at a daily dose of 20, 40 and 100 μg , respectively per kilogram of shrimp's weight. Group 4 received a high molecular weight LPS as admixed with moist pellets at a daily dose of 1000 μg per kilogram of yellowtail's weight. Group 5 (control) received LPS-free moist pellets. On day 7 after supply of feeds, the yellowtails were intraabdominally inoculated with *Enterococcus Seriolicida* as a

pathogen causing enterococcus disease of yellowtail in an amount of 4.0×10^6 cells per yellowtail. A mortality 15 days after inoculation was determined. The results are shown in Tables 9 and 10.

5

Table 9

Group	Days after infection								
	1	2	3	4	5	6	7	8	9
Group 1 low MW LPS 20 μ g/kg	0	0	0	0	0	0	0	0	1*
Group 2 low MW LPS 40 μ g/kg	0	0	0	1	1	2	2	4	4
Group 3 low MW LPS 100 μ g/kg	0	0	0	0	0	1	3	3	5
Group 4 high MW LPS 1000 μ g/kg	0	0	0	1	1	1	3	3	3
Group 5 feed free of LPS	0	0	1	2	7	7	10	12	16

* The number indicates the total number of dead yellowtails.

(Other numbers show the same.)

Table 10

Group	Days after infection						Mortality (%)
	10	11	12	13	14	15	
Group 1 low MW LPS 20 μ g/kg	3	3	3	3	4	4	13.3 ***
Group 2 low MW LPS 40 μ g/kg	7	8	8	8	8	8	26.7 **
Group 3 low MW LPS 100 μ g/kg	5	5	5	7	7	7	23.3 **
Group 4 high MW LPS 1000 μ g/kg	5	9	10	10	11	11	36.7 **
Group 5 feed free of LPS	16	16	17	22	22	22	73.3

** : significant difference between this group and Group 5

($P < 0.05$)

***: significant difference between this group and Group 5
($P < 0.01$)

On 15th day after inoculation of *E. Seriolicida*, 73.3% of
5 yellowtails died in the control group receiving LPS-free feed.
In contrast, a low mortality is indicated by the groups
receiving the low molecular weight LPSSs of the present
invention, i.e. 13.3% from the group receiving 20 μg , 26.7%
from the group receiving 40 μg and 23.3% from the group
10 receiving 100 μg . In other words, there is a significant
difference ($P < 0.05$) in mortality between these groups and the
control group. On the other hand, a mortality of 36.7%
resulted from the group receiving 1000 μg of high molecular
weight LPS. This group showed a higher mortality than the
15 groups receiving low molecular weight LPSSs. The above results
show that the low molecular weight LPSSs of the present
invention can protect fishes against viral infection and are
more efficacious than conventional high molecular weight LPSSs.

20 INDUSTRIAL APPLICABILITY

According to the present invention, there is provided a
safe feedstuff additive for growing crustaceans and fishes, the
feedstuff additive being capable of preventing infectious
diseases by properly activating their intrinsic immune function
25 even when used in a small amount, being capable of preventing
the perish of crustaceans and fishes, and being free from the
problems of public health hazards such as the feedstuff
additive remaining in crustaceans and fishes.

CLAIMS

1. A feedstuff additive for crustaceans or fishes, characterized in that it is prepared from gram-negative bacteria, that it has a molecular weight of 5000 ± 2000 as measured by SDS-PAGE method using a protein marker, that it is substantially free of high molecular weight lipopolysaccharide, that it contains a low molecular weight lipopolysaccharide as an effective component and that it is capable of activating immunity or preventing infection in crustaceans or fishes.

2. A feedstuff additive for crustaceans or fishes comprising the low molecular weight lipopolysaccharide of claim 1 and a carrier acceptable for crustaceans and fishes.

3. Use of the low molecular weight lipopolysaccharide of claim 1 for the preparation of a feedstuff additive for crustaceans or fishes.

4. A method of activating immunity or preventing infection in crustaceans or fishes comprising administering an effective amount of the low molecular weight lipopolysaccharide of claim 1 to crustaceans or fishes.

5. An agent for preventing the perish of crustaceans or fishes comprising the low molecular weight lipopolysaccharide of claim 1 as an effective component.

6. An agent for preventing the perish of crustaceans or fishes comprising the low molecular weight lipopolysaccharide of claim 1 and a carrier acceptable for crustaceans and fishes.

7. Use of the low molecular weight lipopolysaccharide of claim 1 for the preparation of an agent for preventing the

perish of crustaceans or fishes.

8. A method of preventing the perish of crustaceans or fishes comprising administering an effective amount of the low molecular weight lipopolysaccharide of claim 1 to crustaceans or fishes.

9. A feedstuff additive according to claim 1, wherein the gram-negative bacteria are those pertaining to genus Pantoea.

10. A feedstuff additive according to claim 9, wherein the gram-negative bacteria are Pantoea agglomerans.

11. A feed for crustaceans or fishes comprising the feedstuff additive of claim 1.

12. A feed for crustaceans or fishes comprising the agent for preventing the perish of claim 5.

13. A method of breeding crustaceans or fishes comprising administering a feed of claim 11 to crustaceans or fishes.

14. A method of breeding crustaceans or fishes comprising administering a feed of claim 12 to crustaceans or fishes.

15. A feedstuff additive according to claim 1, wherein the infectious diseases is acute viremia of crustaceans, their vibrio diseases, parasitosis or mycosis ; iridovirus infectious diseases of fishes, their rhabdovirus diseases, neuronecrosis, 5 infectious hemopoietic organ necrosis, pseudotuberculosis, streptococcic diseases, enterococcus diseases, vibrio diseases, cold-water disease, Pseudomonas diseases, gliding-bacteria diseases or Saprolegnia diseases.

16. A feedstuff additive according to claim 1, wherein the high molecular weight lipopolysaccharide is one having a molecular weight of at least 8,000.

ABSTRACT OF THE DISCLOSURE

A feedstuff additive for crustaceans and fishes, characterized in that it is prepared from gram-negative bacteria, that it has a molecular weight of 5000 ± 2000 as measured by SDS-PAGE method using a protein marker, that it is substantially free of high molecular weight lipopolysaccharide, that it contains a low molecular weight lipopolysaccharide as an effective component and that it is capable of activating immunity and preventing infection in crustaceans or fishes ; and a feed for crustaceans and fishes comprising the above feedstuff additive.

Declaration For U.S. Patent Application

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled
(Insert Title) ADDITIVES FOR CRUSTACEAN OR FISH FEEDS AND FEEDS

the specification of which is attached hereto unless the following box is checked:

☒ was filed on March 23, 2000 as PCT International Application
 Number PCT/JP00/01764 and was amended on _____
 and/or was filed on _____ as United States Application
 Number _____ and was amended on _____

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claim(s), as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 C.F.R. §1.56.

I hereby claim foreign priority benefits under 35 U.S.C. §119(a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate, or §365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate or PCT International Application having a filing date before that of the application(s) for which priority is claimed:

(List prior foreign applications. See note A on back of this page)	<u>84,399/1999</u> (Number)	<u>Japan</u> (Country)	<u>26/March/1999</u> (Day/Month/Year Filed)	Priority Claimed <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
	<u> </u> (Number)	<u> </u> (Country)	<u> </u> (Day/Month/Year Filed)	<input type="checkbox"/> Yes <input type="checkbox"/> No
	<u> </u> (Number)	<u> </u> (Country)	<u> </u> (Day/Month/Year Filed)	<input type="checkbox"/> Yes <input type="checkbox"/> No

I hereby claim the benefit under 35 U.S.C. §119(e) of any United States provisional application(s) listed below.

<u> </u> (Application Number)	<u> </u> (Filing Date)
<u> </u> (Application Number)	<u> </u> (Filing Date)

(See Note B on back of this page)

☐ See attached list for additional prior foreign or provisional applications.

I hereby claim the benefit under 35 U.S.C. §120 of any United States application(s) or §365(c) of any PCT International application(s) designating the United States of America listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior application(s) (U.S. or PCT) in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 C.F.R. §1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

(List prior U.S. Applications or PCT International applications designating the U.S.)	<u> </u> (Application Serial No.)	<u> </u> (Filing Date)	<u> </u> (Status) (patented, pending, abandoned)
	<u> </u> (Application Serial No.)	<u> </u> (Filing Date)	<u> </u> (Status) (patented, pending, abandoned)

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

(See Note C on back of this page)

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